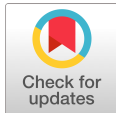


## ARTICLE

# Antimicrobial Action of *Raphanus raphanistrum* subsp. *sativus* (radish) Extracts against Foodborne Bacteria Present in Various Milk Products: A Preliminary Study

Hyun-Woo Lim<sup>1†</sup>, Kwang-Young Song<sup>1,2\*</sup>, Jung-Whan Chon<sup>1,3</sup>, Dongkwan Jeong<sup>4</sup>, and Kun-Ho Seo<sup>1†</sup><sup>1</sup>KU Center for Food Safety and Dept. of Public Health, College of Veterinary Medicine, Konkuk University, Seoul, Korea<sup>2</sup>Dept. of Biological Engineering, Yanbian University of Science and Technology, Yanji, JL, China<sup>3</sup>College of Veterinary Medicine, Oklahoma State University, Stillwater, OK, USA<sup>4</sup>Dept. of Food and Nutrition, Kosin University, Busan, Korea

Received: September 23, 2019

Revised: September 30, 2019

Accepted: September 30, 2019

<sup>†</sup>These authors contributed equally to this study.

\*Corresponding author :  
Kwang-Young Song  
KU Center for Food Safety and Dept. of Public Health, College of Veterinary Medicine, Konkuk University, Seoul, Korea, and Dept. of Biological Engineering, Yanbian University of Science and Technology, Yanji, JL, China.  
Tel : +82-2-450-4121  
Fax : +82-2-3436-4128  
E-mail : drkysong@gmail.com

Copyright © 2019 Korean Society of Milk Science and Biotechnology.

This is an Open Access article distributed under the terms of the Creative Commons Attribution Non-Commercial License (<http://creativecommons.org/licenses/by-nc/3.0>) which permits unrestricted non-commercial use, distribution, and reproduction in any medium, provided the original work is properly cited.

**ORCID**

Hyun-Woo Lim  
<https://orcid.org/0000-0001-6013-8106>  
Kwang-Young Song  
<https://orcid.org/0000-0002-5619-8381>  
Jung-Whan Chon  
<https://orcid.org/0000-0003-0758-6115>  
Dongkwan Jeong  
<https://orcid.org/0000-0002-6305-794X>  
Kun-Ho Seo  
<https://orcid.org/0000-0001-5720-0538>

**Abstract**

Seeds and leaves of *Raphanus raphanistrum* subsp. *sativus* (radish) are known to contain “raphanin,” which has the potential to inhibit pathogenesis associated with foodborne pathogenic bacteria and fungi. In this study, ethanol extracts from *R. raphanistrum* subsp. *sativus* (radish) powder was evaluated for antimicrobial action against 6 different foodborne pathogenic bacteria. The current study demonstrated the potential of *R. raphanistrum* subsp. *sativus* (radish) in inhibiting the growth of *Salmonella* enteritidis 110, *Cronobacter sakazakii* KCTC 2949, *Bacillus cereus* ATCC 10876, and *Staphylococcus aureus* ATCC 6538. However, these antimicrobial action were not observed against *Listeria monocytogenes* ATCC 51776 and *Escherichia coli* 23716. Hence, this study indicates that *R. raphanistrum* subsp. *sativus* (radish) could be used as a natural biopreservative with antimicrobial effects for improving food safety, and as a functional food in the commercial food industry.

**Keywords**

*Raphanus raphanistrum* subsp. *sativus* (radish), antimicrobial action, foodborne pathogenic bacteria, dairy products

**Introduction**

*Raphanus raphanistrum* subsp. *sativus* (radish) belong to a member of Brassicaceae family was a perennial plant and an edible root vegetable [1] (Fig. 1). According to the size, shape, and color of their thick roots of *Raphanus raphanistrum* subsp. *sativus* (radish), there were many different varieties available [1-4]. In general, the most usable and edible part of *Raphanus raphanistrum* subsp. *sativus* (radish) was the root, and also the leaves and stem had been utilized for flavoring or preservation of food [1,5]. The nutritional value of *Raphanus raphanistrum* subsp. *sativus* (radish) was composed of carbohydrates, high amounts of fiber, low amounts of fat, many essential minerals, vitamins, and so on [6]. *Raphanus raphanistrum* subsp. *sativus* (radish) was known to be very important as a medicinal property. Until now, *Raphanus raphanistrum* subsp. *sativus* (radish) was widely exploited around the world as traditional medicine for treatment such as ailments and disorders by various factors which included anemia, female (and/or male) infertility, gastrointestinal systems, respiratory, skin, urinary, and

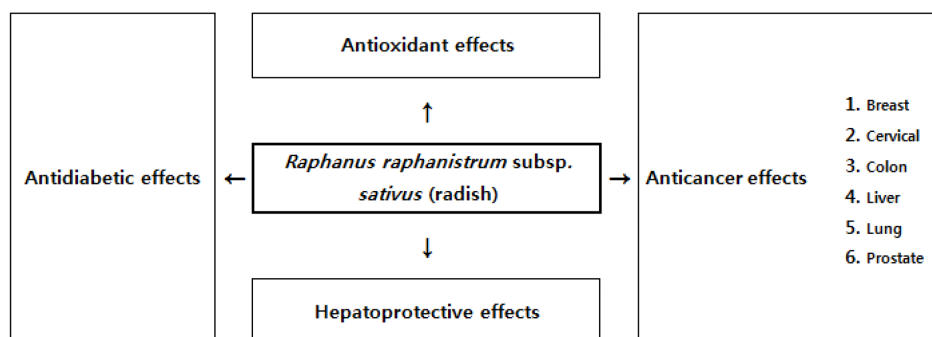


**Fig. 1.** *Raphanus raphanistrum* subsp. *sativus* (radish) belong to the Brassicaceae family and was an edible root vegetable with a pungent taste. The parts of the *Raphanus raphanistrum* subsp. *sativus* (radish) plant that were commonly consumed as the leaves, flowers, pods, and seeds around the world [2,3].

so on [1,7]. According to the report of Manivannan *et al.* *Raphanus raphanistrum* subsp. *sativus* (radish) had been reported to possess (1) antioxidant effects, (2) Hepatoprotective effects, (3) anticancer effects, (4) antidiabetic effects, and so on [8] (Fig. 2).

Especially, the leaves and roots of *Raphanus raphanistrum* subsp. *sativus* (radish) were used as antimicrobial substances [4,9]. It was generally know that the pharmacological characteristics of *Raphanus raphanistrum* subsp. *sativus* (radish) were caused by a wide range of 2<sup>nd</sup> metabolites. Secondary metabolites included alkaloids, antioxidant enzymes, carotenoids, coumarins, flavonoids (including anthocyanins), glucosinolates, phenolics, terpenes, and other compounds [10-13]. Furthermore, the usability of natural extracts has recently increased for improving human's health [14].

In fact, the world's most important issue was foodborne disease. Especially, various foodborne pathogenic bacteria could be easily spread to humans through milk (or dairy



**Fig. 2.** Various nutraceutical potential of *Raphanus raphanistrum* subsp. *sativus* (radish) for human's health [8].



products) that could stimulate readily the outbreak of serious diseases [15]. Since milk (or dairy products) was recognized as the high nutritious food to improve the human's health, milk (or dairy products) could easily become a habitat for foodborne pathogenic bacteria [15,16]. Therefore, attention would be focused on the safety of milk (or dairy products) [17,18]. Among various foodborne pathogenic bacteria, six different foodborne pathogenic bacteria such as *Listeria monocytogenes* ATCC 51776, *Salmonella* Enteritidis 110, *Escherichia coli* 23716, *Cronobacter sakazakii* KCTC 2949, *Bacillus cereus* ATCC 10876, and *Staphylococcus aureus* ATCC 6538 that were often found in milk (or dairy products) were selected and investigated.

Therefore, this study was aimed at investigating the antimicrobial action of *Raphanus raphanistrum* subsp. *sativus* (radish) on *Listeria monocytogenes* ATCC 51776, *Salmonella* Enteritidis 110, *Escherichia coli* 23716, *Cronobacter sakazakii* KCTC 2949, *Bacillus cereus* ATCC 10876, and *Staphylococcus aureus* ATCC 6538 so as to improve the quality of milk (or dairy products) and also to use it as food additives for preventing diabetes.

## Materials and Methods

### 1. *Raphanus raphanistrum* subsp. *sativus* (radish) and ethanol extraction

The powder of *Raphanus raphanistrum* subsp. *sativus* (radish) produced in Korea was purchased from Heungyildang Food (Korea). The dried powder of *Raphanus raphanistrum* subsp. *sativus* (radish) was macerated in 95% ethanol for at least two days (over 48 hours) with occasionally stirring at ambient temperature. And then, the soluble ingredients were concentrated by rotary evaporator at 50°C until dryness, and then the yield was obtained by ethanol extraction type. These stock solutions were filtrated through 0.2 mm Millipore and stored at -20°C before use.

### 2. Foodborne pathogenic bacteria tested in this study

*Listeria monocytogenes* ATCC 51776, *Salmonella* Enteritidis 110, *Escherichia coli* 23716, *Cronobacter sakazakii* KCTC 2949, *Bacillus cereus* ATCC 10876, and *Staphylococcus aureus* ATCC 6538 were obtained from KU Center for Food Safety, College of Veterinary, Konkuk University in Seoul, Korea. These bacteria were grown on nutrient agar (NA) (Oxoid, UK) overnight. Colonies were transferred into tubes containing cryopreservation fluid according to the instruction of the manufacturer (Original Microbiology Bead Storage System, STS, Technical Service Consultants Limited, UK). The beads were stored at -70°C until use.

### 3. Spot-on-lawn assay as antimicrobial action testing

The antibacterial action of *Raphanus raphanistrum* subsp. *sativus* (radish) were tested on *Listeria monocytogenes* ATCC 51776, *Salmonella* Enteritidis 110, *Escherichia coli* 23716, *Cronobacter sakazakii* KCTC 2949, *Bacillus cereus* ATCC 10876, and *Staphylococcus aureus* ATCC 6538 using by the spot-on-lawn assay with some modifications [19]. All test bacteria were cultured on Mueller-Hinton broth (MHB; Difco) and incubated

at  $37^{\circ}\text{C} \pm 0.5$  for one day. The culture broth was diluted using MHB to 0.5 McFarland and spread onto Mueller-Hinton agar (MHA; Difco) using sterilized cotton swabs. A total of negative control (0  $\mu\text{L}$ ),  $1 \times$  (10  $\mu\text{L}$ ),  $2 \times$  (20  $\mu\text{L}$ ), and  $3 \times$  (30  $\mu\text{L}$ ) of *Raphanus raphanistrum* subsp. *sativus* (radish) extract was directly dropped onto the surface of the MHA, respectively. The plates were incubated for one day at  $37 \pm 0.5^{\circ}\text{C}$ , and the inhibition zone was observed.

#### 4. Analysis of statistics

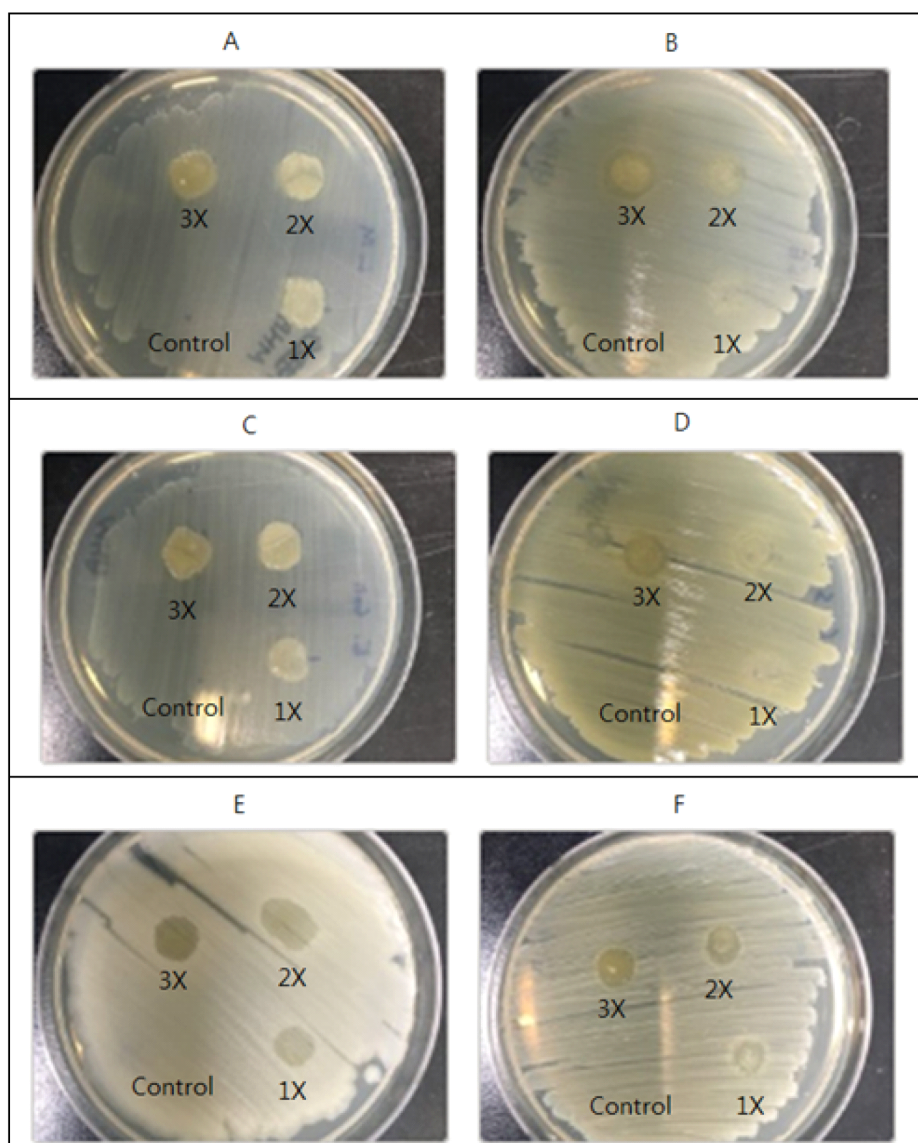
The inhibition of various concentration of *Raphanus raphanistrum* subsp. *sativus* (radish) ethanol extracts against *Listeria monocytogenes* ATCC 51776, *Salmonella* Enteritidis 110, *Escherichia coli* 23716, *Cronobacter sakazakii* KCTC 2949, *Bacillus cereus* ATCC 10876, and *Staphylococcus aureus* ATCC 6538 were analyzed by ANOVA (one-way analysis of variance), and all experiments were carried out independently in duplicate replications, and statistical significance was accepted at the  $p=0.05$  level.

## Results and Discussion

The ethanol extract of *Raphanus raphanistrum* subsp. *sativus* (radish) showed various levels (control,  $1 \times$ ,  $2 \times$ , and  $3 \times$ ) of antimicrobial action when tested by the spot-on-lawn assay (Fig. 3). These results obtained showed that ethanol extract of *Raphanus raphanistrum* subsp. *sativus* (radish) exhibited antimicrobial action against *Salmonella* Enteritidis 110 (partial inhibition), *Cronobacter sakazakii* KCTC 2949 (partial inhibition), *Bacillus cereus* ATCC 10876 (partial inhibition), and *Staphylococcus aureus* ATCC 6538 (total inhibition), and the inhibitory action of *Raphanus raphanistrum* subsp. *sativus* (radish) ethanol extracts was shown as a whole regardless of the increase in the concentration (Fig. 3). Although the final result of antimicrobial action of *Raphanus raphanistrum* subsp. *sativus* (radish) extract by ethanol against *Listeria monocytogenes* ATCC 51776 and *Escherichia coli* 23716 was negative, it did not seem to be effective at all (Fig. 3). In this study, it is likely that the higher concentration, the more effective it will be. Whereas the other two bacteria - *Listeria monocytogenes* ATCC 51776 and *Escherichia coli* 23716- did not show any inhibition by the ethanol extract of *Raphanus raphanistrum* subsp. *sativus* (radish) (Fig. 3).

Until now, it was being used in the treatment of various infection diseases using *Raphanus raphanistrum* subsp. *sativus* (radish) from the perspective of traditional medicine. These attempts have been a major stimulant to study of antimicrobial action of *Raphanus raphanistrum* subsp. *sativus* (radish) [1]. According to various previous researches, it had been shown that *Raphanus raphanistrum* subsp. *sativus* (radish) suppressed the growth of various bacteria, even those that were drug (or antibiotic)-resistant [1]. Also, the root juice of *Raphanus raphanistrum* subsp. *sativus* (radish) could inhibit the growth of *Bacillus subtilis*, *Enterococcus faecalis*, *Escherichia coli*, *Klebsiella pneumoniae*, *Pseudomonas aeruginosa*, *Pseudomonas pyocyaneus*, *Salmonella typhi*, *Staphylococcus aureus*, and so on [20].

Using ethanol, ethylacetate, methanol, petroleum ether and water, crude extracts from



**Fig. 3.** The antimicrobial action on various concentration (control, 1X, 2X, and 3X) of *Raphanus raphanistrum* subsp. *sativus* (radish) ethanol extracts against *Listeria monocytogenes* ATCC 51776 (A), *Salmonella* Enteritidis 110 (B), *Escherichia coli* 23716 (C), *Cronobacter sakazakii* KCTC 2949 (D), *Bacillus cereus* ATCC 10876 (E), and *Staphylococcus aureus* ATCC 6538 (F) tested by the spot-on-lawn assay.

the root-peel of *Raphanus raphanistrum* subsp. *sativus* (radish) black color showed the antimicrobial action against *B. subtilis*, *Bordetella bronchiseptica*, *Enterobacter aerogenes*, *E. coli*, *K. pneumonia*, *Micrococcus luteus*, *P. aeruginosa*, *S. typhi*, *S. aureus*, and so on [21]. Especially, the substances extracted from white and black peel taproots of *Raphanus raphanistrum* subsp. *sativus* (radish) using methanol could have suppress a wide variety of pathogenic bacteria associated with food which were *Arthrobacter atrocyaneus*, *B. sphaericus*, *Corynebacterium ammoniagenes*, *Corynebacterium flavescens*, *Enterobacter hormaechei*, *Kocuria rosea*, *Neisseria subava*, *Pantoea agglomerans*,

*Proteus vulgaris*, *Psychrobacter immobilis*, *Shigella dysenteriae*, and so on [22]. Also, the substances extracted from the root, stem, and leaf of *Raphanus raphanistrum* subsp. *sativus* (radish) white color using chloroform, ethyl acetate and methanol exhibited significantly antimicrobial action against various foodborne and drug-resistant pathogenic bacteria such as *B. subtilis*, clinical isolates of *E. cloacae*, *E. faecalis*, *E. aerogenes*, *E. coli*, *K. pneumonia*, *P. aeruginosa*, *S. epidermidis*, *S. typhimurium*, *S. aureus*, and so on [23]. The substances extracted from seeds of *Raphanus raphanistrum* subsp. *sativus* (radish) white color using ethanol and methanol could strongly show the antimicrobial action against various pathogenic bacteria such as *E. coli*, *K. pneumoniae*, *P. vulgaris*, *P. aeruginosa*, *S. aureus*, *S. sonnei*, *S. typhi*, *S. paratyphi*, and so on [24]. Furthermore, the substances extracted from seeds of *Raphanus raphanistrum* subsp. *sativus* (radish) using methanol and water demonstrated the antimicrobial action against various plant pathogenic bacteria and fungi [25]. The results of this experiment also showed a similar trend to previous studies.

According to Shukla et al. [20], the root juice of *Raphanus raphanistrum* subsp. *sativus* (radish) showed the noticeable antimicrobial action against both gram-positive as *Staphylococcus aureus* and gram-negative as *Enterococcus faecalis*, *Escherichia coli*, *Klebsiella pneumonia* and *Pseudomonas aeruginosa* with the MIC range from 0.078 mg/mL to 0.625 mg/mL. On the other hand, the MIC range of ampicillin as standard antibiotic showed from 0.078 mg/mL to 0.312 mg/mL. The results of this experiment also showed a similar trend to previous studies. According to Jadoun et al. [1], the substances extracted from seeds of *Raphanus raphanistrum* subsp. *sativus* (radish) red color using ethanol demonstrated the antimicrobial action against 5 different pathogenic bacteria such as *E. coli*, *K. pneumonia*, *S. pyogenes*, *S. typhimurium*, and *S. aureus* with the inhibition zone from  $9\pm 2$  mm to  $20\pm 2$  mm. On the other hand, the inhibition zone of chloramphenicol (10 mg/mL) as standard antibiotic exhibited from  $15\pm 1$  mm to  $30\pm 2$  mm [1].

In fact, there were many studies for investigating the antimicrobial action of *Raphanus raphanistrum* subsp. *sativus* (radish). However, very little research had been done on the antimicrobial action of seeds of *Raphanus raphanistrum* subsp. *sativus* (radish) red color [1]. On the contrary, according to other previous studies, the substances extracted from seeds of *Raphanus raphanistrum* subsp. *sativus* (radish) using mixture of dichloromethane and methanol demonstrated the lack of antimicrobial action against 14 different bacterial and fungal strains [26]. But, Jadoun et al. addressed that crude substances extracted from seed of *Raphanus raphanistrum* subsp. *sativus* (radish) was considerably effective against all pathogenic bacteria tested [1]. It was thought that the different findings between the two studies may be due to reason of (1) the extraction methods and (or) the solvents used [22,24,27].

Especially, according to report of Jamuna et al. [27], the cold extraction of fresh root of *Raphanus raphanistrum* subsp. *sativus* (radish) was significantly better than dried cold extraction or soxhlet in antioxidant activities tested. Also, according to study of Ahmad et al. [24], the antimicrobial action against 8 different pathogenic bacteria including gram-positive and gram-negative was better in extracts of seed of *Raphanus rapha-*





*nistrum* subsp. *sativus* (radish) white color using ethanol and methanol than in acetate, benzene, chloroform, ethyl acetate, and water. Furthermore, Janjua et al. reported there were differences in antimicrobial action depending on the solvent used such as ethanol, ethyl acetate or others in peels of *Raphanus raphanistrum* subsp. *sativus* (radish) black color [21]. One thing to be careful about was that the antimicrobial action of the substances extracted from various solvents varied widely, especially depending on the species of bacteria and on genetic variation [21].

Therefore, the potential antimicrobial action by the unknown plant-derived substance became increasingly interested so as to prevent the outbreak of various drug-resistant pathogenic bacteria [28]. Especially, the substances extracted from various plants should be widely examined for its possibility for natural biopreservatives such as antimicrobial action, since many consumers urgently needed for natural preservatives for substituting artificially manufactured chemicals [1,6]. Also, research on various substances extracted from various *Raphanus raphanistrum* subsp. *sativus* (radish) produced in each country would have to be carried out.

In conclusion, this study demonstrated the potential of *Raphanus raphanistrum* subsp. *sativus* (radish) to inhibit the growth of *Salmonella* Enteritidis 110, *Cronobacter sakazakii* KCTC 2949, *Bacillus cereus* ATCC 10876, and *Staphylococcus aureus* ATCC 6538 as antimicrobial action, except for *Listeria monocytogenes* ATCC 51776 and *Escherichia coli* 23716. Hence, this study indicated that *Raphanus raphanistrum* subsp. *sativus* (radish) could be applicable in natural biopreservatives such as antimicrobial action for improving the food safety, and also in functional food material of the food industry.

## Conflict of Interest

The authors declare no potential conflict of interest.

## Acknowledgements

This research was supported by Basic Science Research Program through the National Research Foundation of Korea (NRF) funded by the Ministry of Education (NRF-2017R1D1A1B03035427).

## References

1. Jadoun J, Yazbak A, Rushrush S, Rudy A, Azaizeh H. Identification of a new antibacterial sulfur compound from *Raphanus sativus* seeds. *Evid-Based Compl Alt*. 2016;2016:9271285.
2. USDA. 2009. Radishes: a new cover crop option. *Crops Soils*. 14-17.
3. Nagdeve M. 20 Surprising benefits of radish [Internet]. 2019 [cited 2019 Aug 25]. *Organic fact*. Available from: <https://www.organicfacts.net/health-benefits/vegetable/health-benefits-of-radish.html>
4. Aruna G, Yerragunt VG, Raju AB. Photochemistry and pharmacology of *Raphanus*

- sativus*. Int J Drug Formul Res. 2012;3:43-52.
5. Beevi SS, Mangamoori LN, Dhand V, Ramakrishna DS. Isothiocyanate profile and selective antibacterial activity of root, stem, and leaf extracts derived from *Raphanus sativus* L. Foodborne Pathog Dis. 2009;6:129-136.
  6. Manchali S, Chidambara Murthy KN, Patil BS. Crucial facts about health benefits of popular cruciferous vegetables. J Funct Foods 2012;4:94-106.
  7. Said O, Khalil K, Fulder S, Azaizeh H. Ethnopharmacological survey of medicinal herbs in Israel, the Golan heights and the West Bank region. J Ethnopharmacol. 2002;83:251-265.
  8. Manivannan A, Kim JH, Kim DS, Lee ES, Lee HE. Deciphering the nutraceutical potential of *Raphanus sativus*: a comprehensive overview. Nutrients 2019;11:402.
  9. Gutierrez RM, Perez RL. *Raphanus sativus* (Radish): their chemistry and biology. Sci World J. 2004;4:811-837.
  10. Fahey JW, Zalcmann AT, Talalay P. The chemical diversity and distribution of glucosinolates and isothiocyanates among plants. Phytochemistry 2001;56:5-51.
  11. Montaut S, Barillari J, Iori R, Rollin P. Glucoraphasatin: chemistry, occurrence, and biological properties. Phytochemistry 2010;71:6-12.
  12. Kim KH, Kim CS, Park YJ, Moon E, Choi SU, Lee JH, et al. Anti-inflammatory and antitumor phenylpropanoid sucrosides from the seeds of *Raphanus sativus*. Bioorg Med Chem Lett. 2015;25:96-99.
  13. Shin T, Ahn M, Kim GO, Park SU. Biological activity of various radish species. Orient Pharm Exp Med. 2015;15:105-111.
  14. Jurikova T, Mlcek J, Skrovankova S, Sumczynski D, Sochor J, Hlavacova I, et al. Fruits of black chokeberry *Aronia melanocarpa* in the prevention of chronic diseases. Molecules 2017;22:E944.
  15. Zeinhom MMA, Abdel-Latef GK. Public health risk of some milk borne pathogens. Beni-Suef Univ J Basic Appl Sci. 2014;3:209-215.
  16. El-Baz AH, El-Sherbini M, Abdelkhalek A, Al-Ashmawy MA. Prevalence and molecular characterization of *Salmonella serovars* in milk and cheese in Mansoura city, Egypt. J Adv Vet Anim Res. 2017;4:45-51.
  17. Nicklas TA. Calcium intake trends and health consequences from childhood through adulthood. J Am Coll Nutr. 2003;22:340-356.
  18. Boor KJ, Wiedmann M, Murphy S, Alcaine S. A 100-year review: microbiology and safety of milk handling. J Dairy Sci. 2017;100:9933-9951.
  19. Cadirci BH, Citak S. A comparison of two methods used for measuring antagonistic activity of lactic acid bacteria. Pakistan J Nutr. 2005;4:237-241.
  20. Shukla S, Chatterji S, Yadav DK, Watal G. Antimicrobial efficacy of *Raphanus sativus* root juice. Int J Pharm Bio Sci. 2011;3:89-92.
  21. Janjua S, Shahid M, Abbas FI. 2013. Phytochemical analysis and *in vitro* antibacterial activity of root peel extract of *Raphanus sativus* L. var. *niger*. Adv Med Plant Res. 2013;1:1-7.
  22. Umamaheswari M, Ajith MP, Asokkumar K, Sivashanmugam T, Subhadradevi V, Jagannath P, Madeswaran A. *In vitro* angiotensin converting enzyme inhibitory and





- antioxidant activities of seed extract of *Raphanus sativus* Linn. Cent Eur J Exp Biol. 2012;1:11-17.
23. Shrivastava N, Bhargava R. Antibacterial potential of selected medicinal plants against *Escherichia coli* and *Staphylococcus aureus*. Acta Biologica Indica. 2012;1: 133-135.
  24. Ahmad F, Hasan I, Chishti DK, Ahmad H. Antibacterial activity of *Raphanus sativus* Linn. Seed extract. Global J Med Res. 2012;12:25-33.
  25. Rani I, Akhund S, Abro H. 2008. Antimicrobial potential of seed extract of *Raphanus sativus*. Pak J Bot. 2008;40:1793-1798.
  26. NCCLS [National Committee for Clinical Laboratory Standards]. 2000. Approved standard M7-A5: methods for dilution antimicrobial susceptibility test for bacteria that grow aerobically. 5th ed. Clinical and Laboratory Standards Institute.
  27. Jamuna KS, Ramesh CK, Mahmood R, Pallavi M, Aditya Rao SJ. Effect of different extraction methods on total phenolic content and antioxidant activities of *Raphanus sativus*. Int J Biol. 2015;4:4653-4657.
  28. Liepina I, NikoLajeva V, Jakobsone I. Antimicrobial activity of extracts from fruits of *Aronia melanocarpa* and *Sorbus aucuparia*. Environ Exp Biol. 2013;11:195-199.